

Introduction

FTMS instruments are inherently complex and require FTMS fundamentals understanding to unlock their full capability and avoid results misinterpretation. Here we present the FTMS Teaching Pack - the package which includes the FTMS fundamental tutorials, FTMS Simulator software, and customized training case studies.



FTMS SIMULATOR

FTMS Simulator is a software tool to accurately simulate FTMS isotopic envelopes and mass spectra:

- to visually validate and verify the experimental results and hypotheses, perform data quality control
- to teach the FTMS subject by demonstrating realistic transients and corresponding mass spectra
- to search the experimental data with a like-for-like simulated data using similarity score's ranking • to design FTMS experiments by selecting optimum settings and FT data processing parameters

Key features

 Analyte parameters selection: elemental composition, amino acid sequence, charge carrier, modifications isotopic enrichment/depletion, number of charge states, and number of isotopologues

• FTMS experiment parameters: FTMS instrument selection (models of Orbitrap[™] & ICR), resolution setting (@ m/z), add noise, set signal intensity thresholds, set initial phase, sampling rate, and decay rate

• Data processing parameters: absorption or magnitude FT, unapodized or apodization function, number of zero-fills, full or reduced profile mass spectra, centroids

• Visualize transients with and without apodization, visualize frequency spectra

• Use mass spectra: manually or automatically set peak intensities, visualize isotopic envelopes, plot multiple isotopic envelopes for a broadband view, extract high quality figures or data points for analysis • Results output: use simulated data to search experimental data, rank search results by the similarity score (m/z), intensity), save project for future use, create report as a PDF file



Figure 1. Schematic diagram of the workflow of Fourier transform (FT) mass spectra simulation for user-defined chemical compounds via in silico time-domain signal (transient) generation for ICR or Orbitrap FTMS instruments. (a) Definition of molecular formulas of interest, (b) calculation of a complete isotopic distribution on a m/z scale, (c) conversion of the calculated isotopic distribution from a m/z scale into a frequency scale, (d) simulation of transients composed of sinusoidal components that correspond to individual isotopologues (m/z values and abundances), (e) FT processing of transients, and (f) visual representation of transients, isotopic distributions, and mass (or frequency) spectra. (JASMS, 2020, 1927; DOI: 10.2021/jasms.0c00190)

Settings

Isotopologues: how many and which ones

FT processing settings

Compound definition:	FT processing settings:			
 Elemental composition 	• FTMS instrument type and model: ICR/MRMS, Orbitraps			
 Amino acid sequence 	• Harmonics order: which harmonic to calculate (1, 2, 3,)			
 Isotopes relative abundances 	Resolution: at target peak, instrument setting, transient length			
 Modifications 	• FT mode: absorption or magnitude			
 Mass (<i>m/z</i>) or frequency value 	Apodization window: none, full (Kaiser), half (semi Kaiser)			
Ion (charged compound) definition:	• Number of zero fills: 0, 1, 2, or 3			
Charge carrier:	• Sampling rate (digitization frequency): 1, 2, 4, 6 MHz, or any			
electron, H ⁺ , K ⁺ , Na ⁺ , Cs ⁺ , I ⁻ , HCOO ⁻	Noise (added to the transient): noise amplitude			
 Ionization: positive, negative, or neutral 	• Decay rate: ion signal decay rate in a transient, e ^{-(decay rate)}			
 Charge state: selected or range 	Phase: initial phase (angle) of ion detection in a transient			

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Apodization

With apodization



— Simulated centroids < •</p> -- Centroids threshold 🗸 🗖 1326 1327 1328 0.00 0.02 0.04 0.06 0.08 0.10 0.12 0.14 0.16 0.18 0.20 0.22 0.24 Time s 1326 1327 1328 1329 Figure 5. The apodization windows "shape" transients prior to Fourier Figure 6. Comparison of mass spectra processing without (left panel) and transform processing. (Left panel) shows the unapodized transient of a with (right panel) apodization (number of zero-fills in both cases is 2). peptide [PHILADELPHIA+H]⁺ analyzed in Figure 2. (Right panel) shows Expectedly, apodization significantly reduces the FT artefacts and Typically, 2 zero-fills are applied to process modern FTMS data. this time-domain transient after half-window (semi Kaiser) characteristics sidelobes, such as Gibbs oscillations. The associated apodization. reduction in resolution can be also noted.



representation.

FTMS Teaching Pack: an interactive resource to teach FTMS fundamentals

Olga Vvedenskaya, Anton N. Kozhinov, Konstantin O. Nagornov, and Yury O. Tsybin

olya@spectroswiss.ch

Spectroswiss, EPFL Innovation Park, 1015 Lausanne, Switzerland

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Without apodization

- Simulated centroids -

Centroids threshold -



experimental transients and provide a realistic mass spectra

Noise



FTMSSIMULATOR

FT mode / apodization: absorption

With apodization

✓ Check all

— Simulated profile ✓

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Zero-fills / sample rate:

Frequency and resolution

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Figure 3. FTMS simulator allows the user to specify a particular model of an Orbitrap or an ICR FTMS instrument. Both commercially available instruments and those custom developed, e.g., 21 T FT-ICR MS instruments, are included. Once a specific instrument is selected, simulations will take into account period (ior detection time) setting via selection of a resolution target - the typically employed settings are provided to the user. Moreover, the ion detection frequency can be specified at the fundamental harmonic as wel as on the higher order harmonics specified by the user. The frequency spectrum can also be visualized.





Figure 4. Broadband (wide *m/z* range) mass spectra of practically any complexity can be simulated for the user-selected parameters. The summed mass spectra can be extracted as high quality figures or as data points. The latter option shows utility in quality control applications and experimental data search The relative intensity of peaks in broadband mass spectra are user-defined. Top-down mass spectra for protein analysis is a particularly interesting application of this capability. The thus generated data can be used to benchmark and validate software tools, and help in optimizing the processing settings of these tools.









Figure 8. Peak shape changes depending on the number of zero-fills. Higher numbers of zero-fills increase the number of points that define mass





Figure 12. The BioPharma views.





- Supporting protein analysis experiments, including large proteins, such as monoclonal antibodies and viruses
- Enhancing confidence of metabolite analysis with accurate representation of the isotopic fine structure
- Assisting in method development for analysis of complex ADCs, including verification of their critical quality attributes, such as DARs
- Fundamentals understanding and new knowledge creation, for example via analysis of the time-domain isotopic beat patterns
- FTMS Simulator is a foundation for the FTMS Teaching Pack
- FTMS Simulator is capable of simulating FTMS data for any current FTMS application, for example for DAR analysis of complex ADC species
- It can be used to design experiments and support data analysis in FTMS, including for benchmarking data analysis software tools
- Simulated isotopic profiles and centroids can be used to search the experimental data and rank the results using the similarity scores
- The accurately simulated results can reveal FT processing-related artefacts or characteristic features, which are lost in typical calculators
- The didactic nature of the FTMS Simulator appears extremely useful in educating and training the rigorous FTMS scientists
- Didactic materials include the FTMS fundamentals tutorial and case studies, which can be customized or adapted by the user





tool facilitates definition of structural information (elemental composition, sequence, and modifications). mass spectra of the thus defined species can be visualized with the userdefined relative abundances in narrowband and broadband mass spectra



Figure 13. The BioPharma tool can support structural analysis monoclonal antibodies (mAbs)-derived species, including antibody-drug conjugates (ADCs). For example, it can aid in the validation and development of novel methods for the analysis of drug-to-antibody ratios (DARs) of ADCs (Nagornov et al., Anal. Chem. 2021, 12930)

Validation

Appropriate use of the simulated FTMS data in the quality control and method development applications requires careful validation of the employed tool, FTMS Simulator. Therefore the validation procedures have been performed on diverse molecular sizes and complexity levels.

> Figure 14. Benchmarking FTMS Simulator: comparison of the simulated (top panel) and experimental (bottom panel) data. Results are shown for the analysis of trastuzumab (including its proteoforms) performed with a Q Exactive HF Orbitrap FTMS in the denaturing mode (electrospray ionization). The expanded view of the 56 charge state shows the resolved proteoforms, but unresolved isotopic envelopes (64 ms transients). Intensities of the simulated peaks are assigned by the user.

Figure 15. Analysis of peptide isotopic fine structure. Expanded views into (black color, solid line) simulated and (blue color, dashed line) experimental profile mass spectra showing the A+2 isotopologues of a singly charged MRFA peptide. Peak centroids are shown (blue color, dashed line) for experimental and (black color, solid lines) for simulated peaks. Calculated IFS distributions are shown with red color solid lines. Experimental data were acquired using a Q Exactive HF FTMS with 10 m/z isolation (SIM mode); simulated data were obtained for corresponding settings. Both simulated and experimental transients of different length, Tacq = 160 ms (left column), Tacq = 590 ms (middle column), and Tacq = 1000 ms (right column), were apodized using the Kaiser function and FT processed via (top panel) mFT and full window, (middle panel) aFT and full window, and (bottom panel) aFT and half window. (JASMS 2021, 1927; DOI: 10.2021/jasms.0c00190)

Figure 16. Analysis of a 5 kDa protein isotopic distribution. Expanded views into (black color, solid line) simulated and (blue color, dashed line) experimental profile mass spectra showing isotopic distribution of 5+ insulin protein. Peak centroids are shown for experimental peaks (blue color, dashed line) and for simulated peaks (black color, solid lines). Theoretical isotopic pattern is shown with red color solid lines. Experimental data acquired using a Q Exactive HF FTMS, simulated data obtained using the corresponding settings. Both simulated and experimental transients of different length from left: Tacq=10 ms, Tacq=30 ms, Tacq=45 ms and Tacq=75 ms were apodized using Kaiser function and FT processed via (top panel) mFT, full window, (middle panel) aFT, full window, and (bottom panel) aFT, half window. (*JASMS* 2021, 1927; DOI: 10.2021/jasms.0c00190)

Case studies

Several case studies and online tests have been prepared by Spectroswiss team. Further developments can be easily made due to the flexibility of the FTMS teaching pack that allows for the custom tasks creation including but not limited to:

• Creating generic FTMS working problems for Orbitraps and ICR/MRMSs instruments (resolution change, transient parameters, frequencies, signal amplitudes, signal-to-noise ratio calculations) for various data representation types (mFT, aFT)

Conclusions

• It is a powerful and versatile tool created for simulation of realistic FTMS data via transient generation